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## Introduction

- Klebsiella pneumoniae* is a gram-negative pathogen associated with multidrug resistance.
- Colistin is one of the last resorts antibiotics for treatment of multidrug-resistant *K. pneumoniae* infections [1].
- Emergence of colistin resistance is a global issue [2] and can develop during colistin treatment [3].
- Quantitative modelling of time-kill experiments can guide the design of treatment schedules to control the emergence of resistance [4].

## Aims

- Characterize the effect of colistin on kill and growth kinetics of *K. pneumoniae* using a static time-kill (STK) experiment and population analysis profiling (PAP).
- Develop a quantitative model to characterize the dynamics of colistin resistance during treatment based on STK and PAP data.

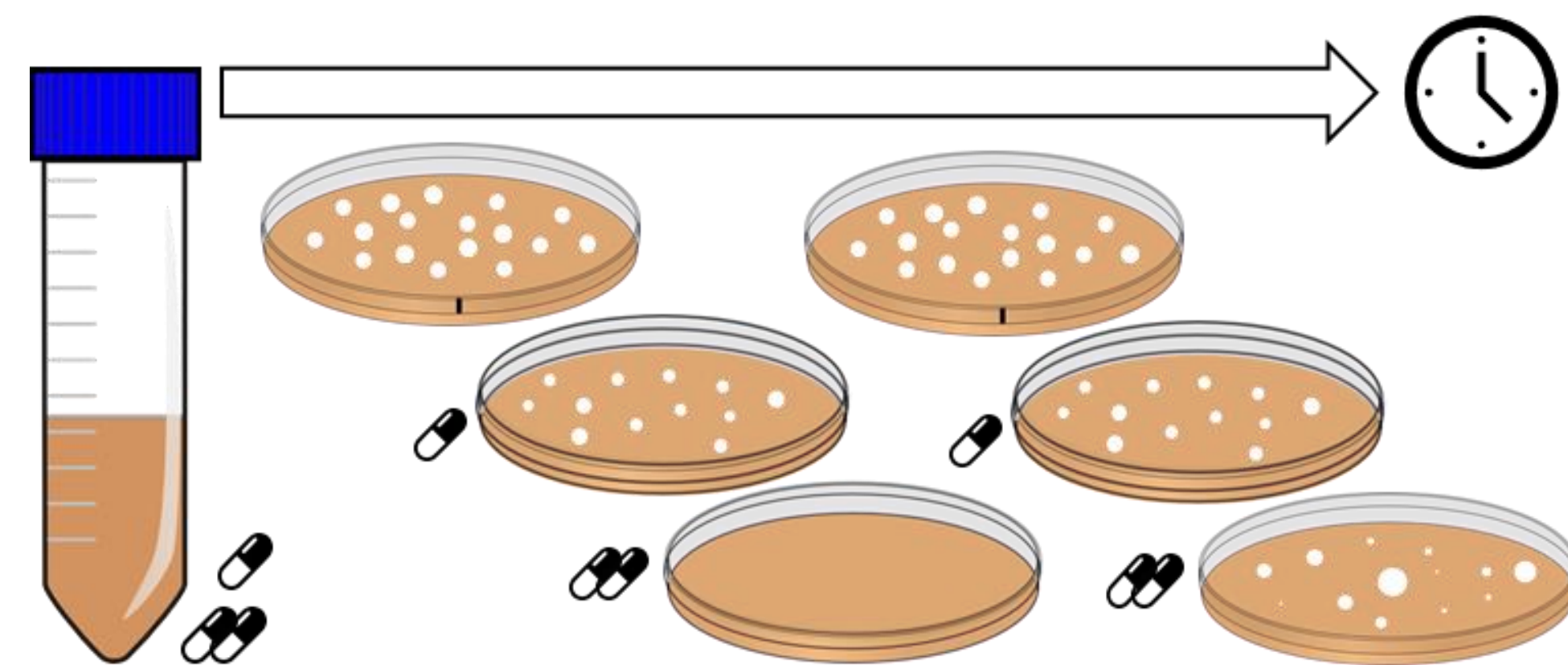
## Methods

### Experimental design

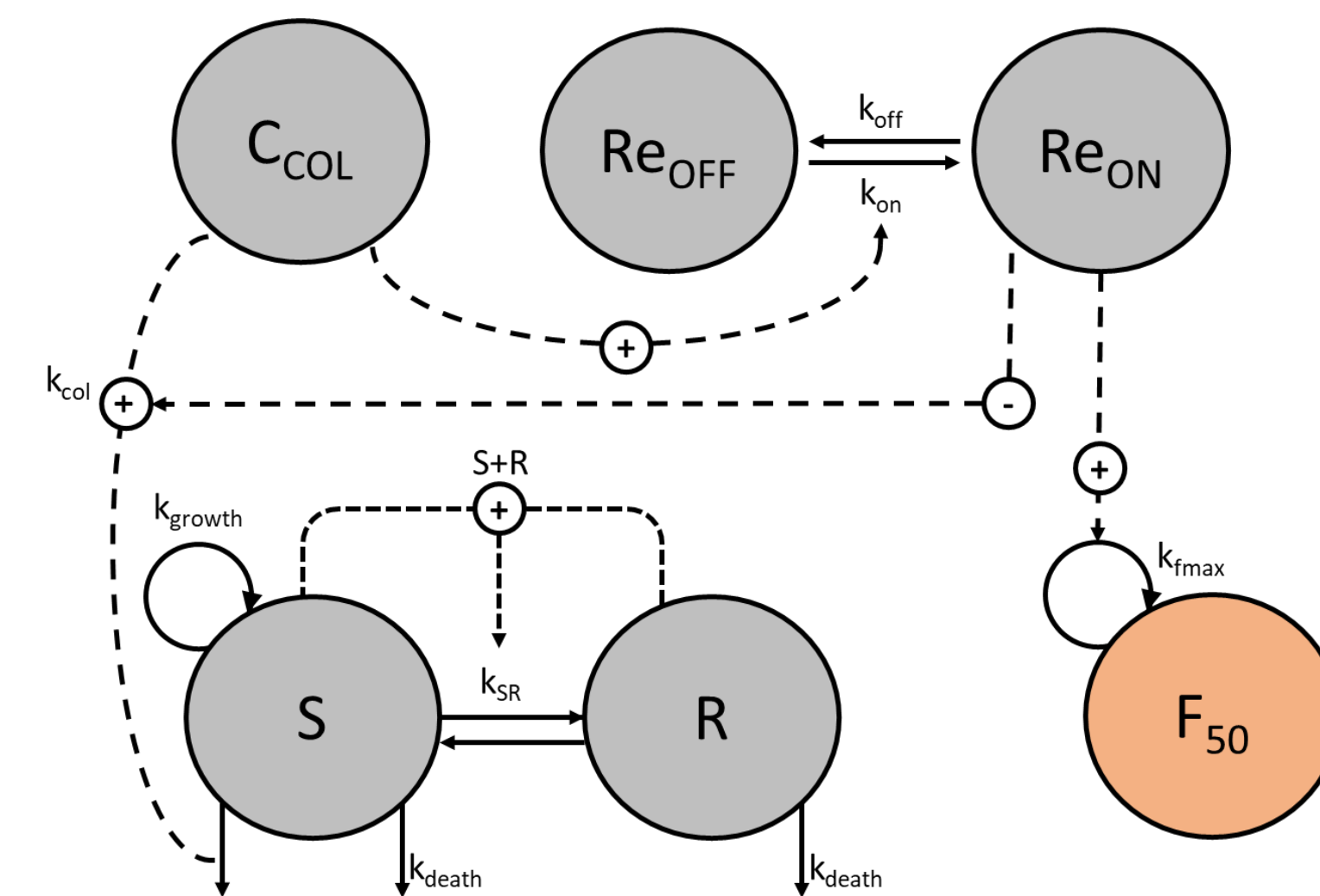
- STK experiments were carried out by exposing the carbapenemase-producing *K. pneumoniae* ATCC BAA-1705 to different colistin concentrations.
- Colony forming units (CFU) were determined at 0, 1, 2, 4, 8, and 24 h for antibiotic free and PAP-plates with increasing colistin concentrations.

### Model development

- A previously developed pharmacodynamic model for colistin time-kill kinetics was used as base model [4], including:
  - drug-susceptible growing bacteria (S) and non-susceptible bacteria (R);
  - drug concentration ( $C_{col}$ )-dependent rate influencing the transition from the state of no resistance ( $Re_{OFF}$ ) to resistance ( $Re_{ON}$ );
  - concentration-effect relationship for bacterial kill and resistance development according to an Emax model.
- The PAP data was modelled with an extension from the base model based on the ratio of colistin resistance and total CFU counts.



**Figure 1. Overview of time kill experiment.** Total CFU sampled at a given time point are quantified on multiple PAP plates which contain colistin concentration ranged from 0 to 20mg/L

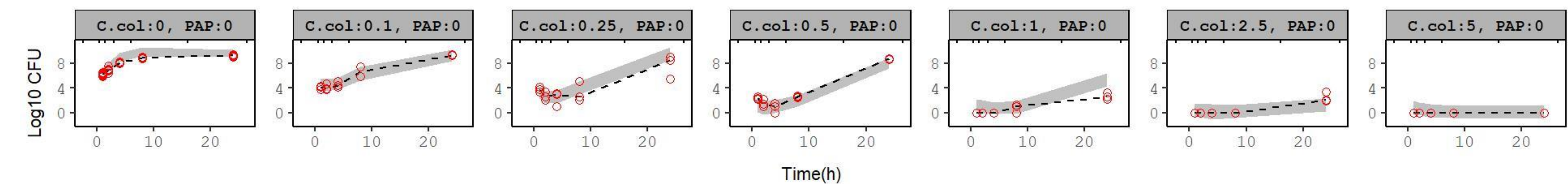


**Figure 2. Schematic structure of the developed PD model.** The grey compartment represents the existing model. We added a compartment for the F50 to incorporate PAP data.

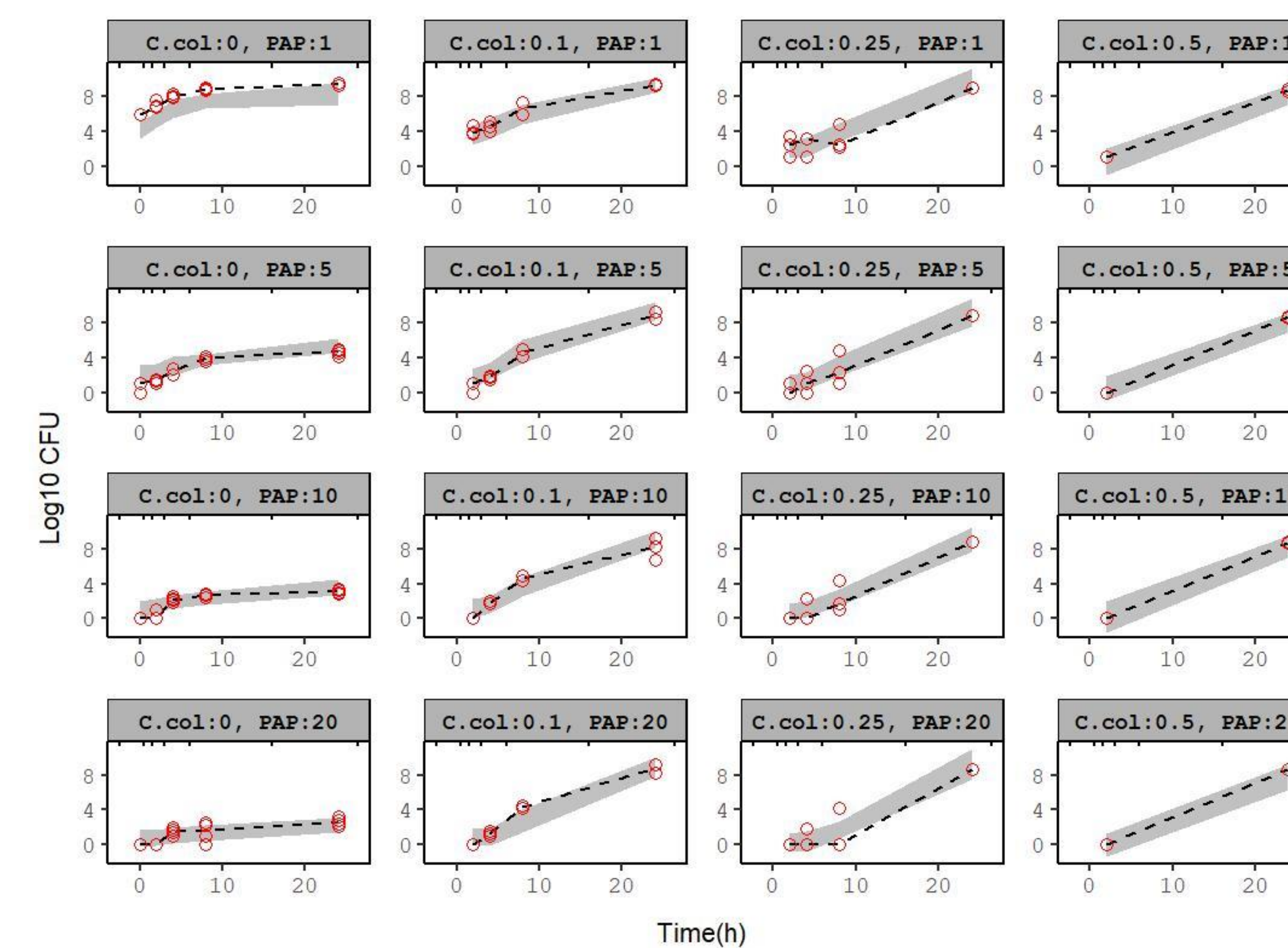
## Results

- Bacteria at the baseline were assumed to contain only sub-population S.
- The bacteria fraction (F) survived at specific concentrations of colistin PAP-plates ( $C_{colPAP}$ ) was described in eq. 1-2.
 
$$\log_{10}(CFU_{PAP}) = F * \log_{10}(CFU_{total}) \quad (1)$$

$$F = 1 - (C_{colPAP} / (F_{50}(t) + C_{colPAP})) \quad (2)$$
- The emergence of resistance toward antibiotic on PAP plates described by an Emax equation with  $F_{50}$  at which half of the total population will survive at time t and PAP plate concentration ( $C_{colPAP}$ ).  $F_{50}$  is a time-varying variable (Eq. 3)
 
$$d/dt(F_{50}) = k_{fmax} * Re_{ON} * \beta * F_{50} \quad \text{with } F_{50}(t=0) = EC_{50} \quad (3)$$
- $k_{fmax}$  reflects the maximum emergence rate of  $F_{50}$ .
- The model adequately captured total CFU time-kill profiles and PAP CFU data.



**Figure 3. CFU observations and model predictions for the STK experiment.** Grey areas indicate 95% model prediction interval of median (colistin concentration from left to right: 0, 0.1, 0.25, 0.5, 1, 2.5, 5mg/)



**Figure 4. CFU observations and model predictions for PAP plates of the STK experiment.** Grey areas indicate 95% model prediction interval of median model predictions (PAP colistin concentration top to bottom: 1, 5, 10, 20 mg/L)

**Table 1. Population parameter estimates for the PD model.**

Parameter (unit)	Description	Estimate	RSE%
$k_{growth}$ ( $h^{-1}$ )	Bacterial growth rate	1.9	16.1
$k_{death}$ ( $h^{-1}$ )	Natural bacterial death rate	0.179	fixed
$B_{max}$ ( $\log_{10}$ CFU/mL)	Maximum bacterial population	9.6	1.79
$E_{max}$ ( $h^{-1}$ )	Maximum kill rate	378	4.81
$EC_{50}$ (mg/L)	Concentration that results in half of $E_{max}$	2.36	7.96
$k_{onmax}$ (L/mgh)	Maximal resistance rate in the presence of colistin	3.71	7.08
$C_{on50}$ (mg/L)	Antibiotic concentration that results in half of $k_{onmax}$	2.23	17.8
$\delta$	Hill factor for drug concentration influencing rate constant for the development of adaptive resistance	0.748	8.88
$\beta$	Hill factor for drug inhibition due to resistance	0.158	20.3
$k_{off}$ ( $h^{-1}$ )	Rate constant for bacteria to return to the susceptible state after developing resistance	0.0791	32.8
$k_{fmax}$ ( $h^{-1}$ )	Maximum rate of resistance development toward colistin on PAP plate	0.237	26.8
RES	Additive residual error ( $\log_{10}$ scale)	0.964	

## Conclusion

- Emergence of colistin resistance occurs at low concentration of 0.1mg/L in STK by PAP analysis.
- Our modelling approach based on time-kill studies and PAP assays, is relevant to characterize the rate and extent of resistance.
- The model can be used for simulations of the responses and the changes in (hetero-) resistance profile of *K. pneumoniae* population during colistin treatment.