

# MODELLING PHARMACOGENETIC DATA IN POPULATION STUDIES DURING DRUG DEVELOPMENT

**ADRIEN TESSIER<sup>[1,2]</sup>, JULIE BERTRAND<sup>[3]</sup>, MARYLORE CHENEL<sup>[2]</sup>, EMMANUELLE COMETS<sup>[1,4]</sup>**

[1] IAME, INSERM UMR 1137, University Paris Diderot, Sorbonne Paris Cité

[2] Institut de Recherches Internationales Servier

[3] Genetic Institute, University College of London

[4] INSERM CIC 1414, University Rennes 1

**INTRODUCTION**

---

**CONTEXT**

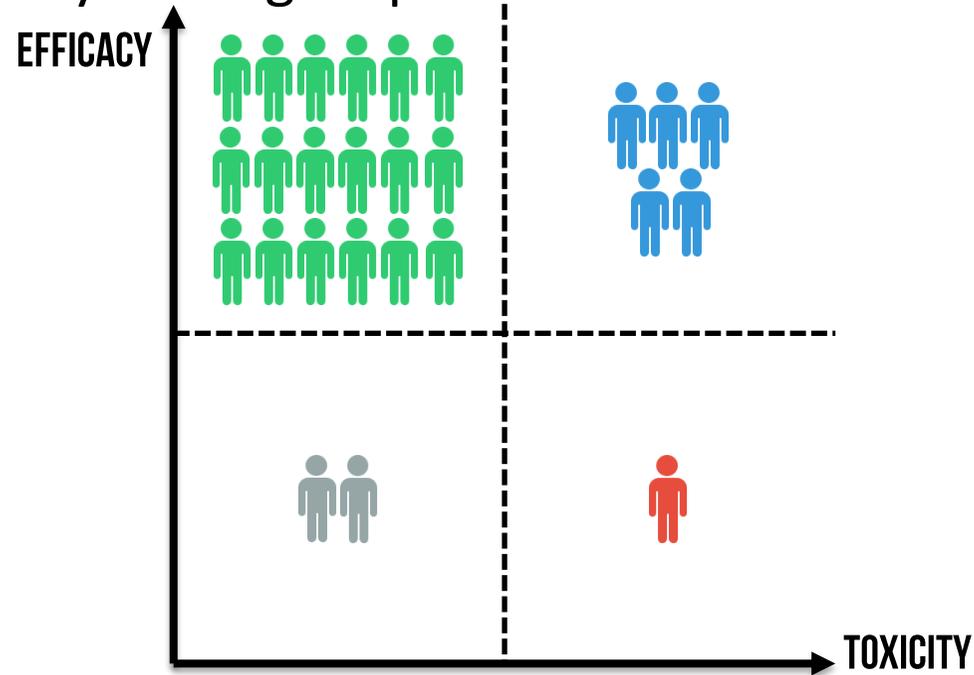
**OBJECTIVES**



# INTRODUCTION: CONTEXT

## PHARMACOGENETICS

**Pharmacogenetics:** study of the variation in genetics in relation to the interindividual variability in drug response<sup>1</sup>



Identifying the genetic variants related to response variability can significantly **increase drug efficacy**<sup>2</sup> and **reduce toxicity**<sup>3</sup>

1. Motulsky 1969      3. Mallal et al. 2008 (abacavir)

2. Rosell et al. 2009 (gefitinib)

# INTRODUCTION: CONTEXT

# PHARMACOGENETICS

**Pharmacogenetics:** study of the variation in genetics in relation to the interindividual variability in drug response<sup>1</sup>

**genetic variation:** Single Nucleotide Polymorphisms (**SNPs**)  
variation of only one base in genomic sequence  
increasingly screened in clinical studies (DNA microarray)

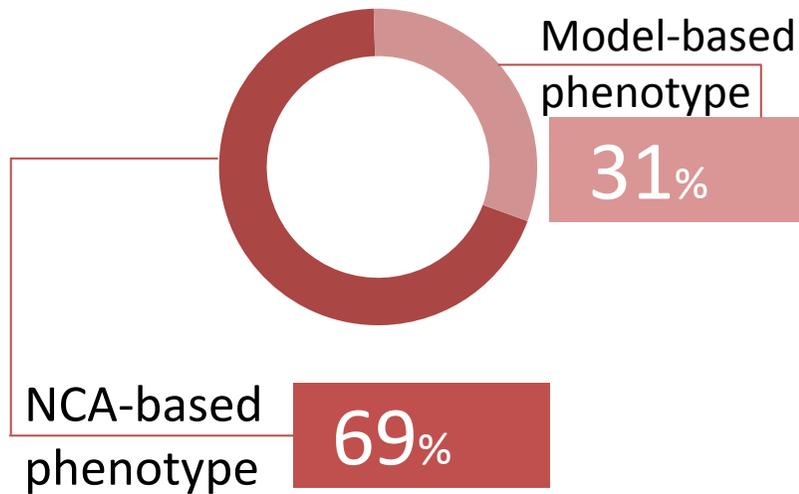
 **TCGATGCATCGG****T****A**TATCTGA  
 **TCGATGCATCGG****C****A**TATCTGA

**interindividual variability in pharmacokinetics (PK):** variation in the enzymatic activity (metabolism, transport)

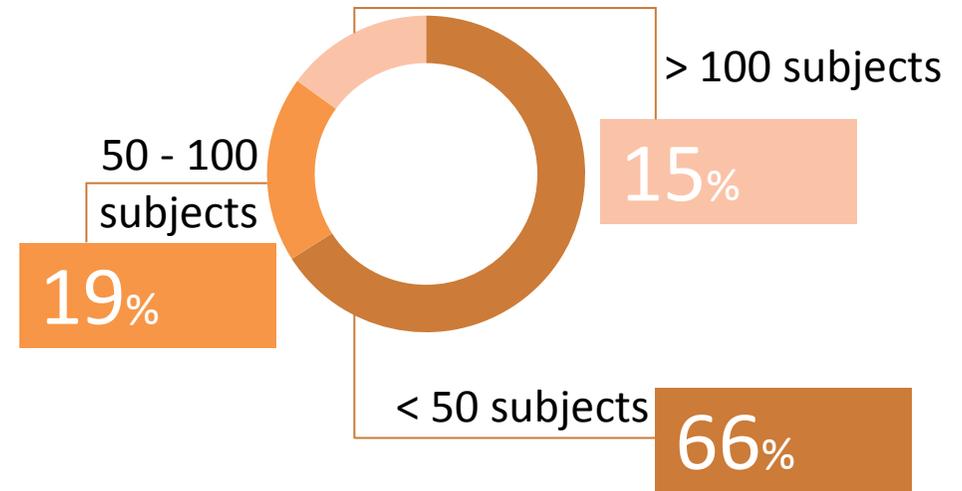
1. Motulsky 1969

# MULTIPLE METHODOLOGIES FOR PHARMACOGENETIC STUDIES

## Phenotype



## Sample size



Survey on 85 pharmacogenetic studies published during the period 2010 - 2012 (Tessier et al. 2015)

Most analyses use NCA-based phenotype estimated from a limited number of subjects

# INTRODUCTION: CONTEXT

# MOTIVATING EXAMPLE

# PHASE I CLINICAL STUDY

A drug S developed by Servier in phase I clinical development

## **PK:**

**N = 78 healthy volunteers**

rich sampling design (**n = 16 samples per subject**)

nonlinear PK with doses

## **Pharmacogenetics:**

all subjects genotyped for **176 SNPs** of genes known to be involved in the PK of drugs (metabolic enzymes, transporters, nuclear receptors)

- **challenging settings to compare NCA and model-based analysis**

# INTRODUCTION: OBJECTIVES

## THREE OBJECTIVES

**PART 1:** Assessment of pharmacogenetic analysis methods

To compare the ability of different **PK phenotypes** to detect genetic effects

To assess the performance of different **association tests**

**PART 2:** Enhance detection of genetic variants through combined designs

To assess **combined analysis** of phase I and II data

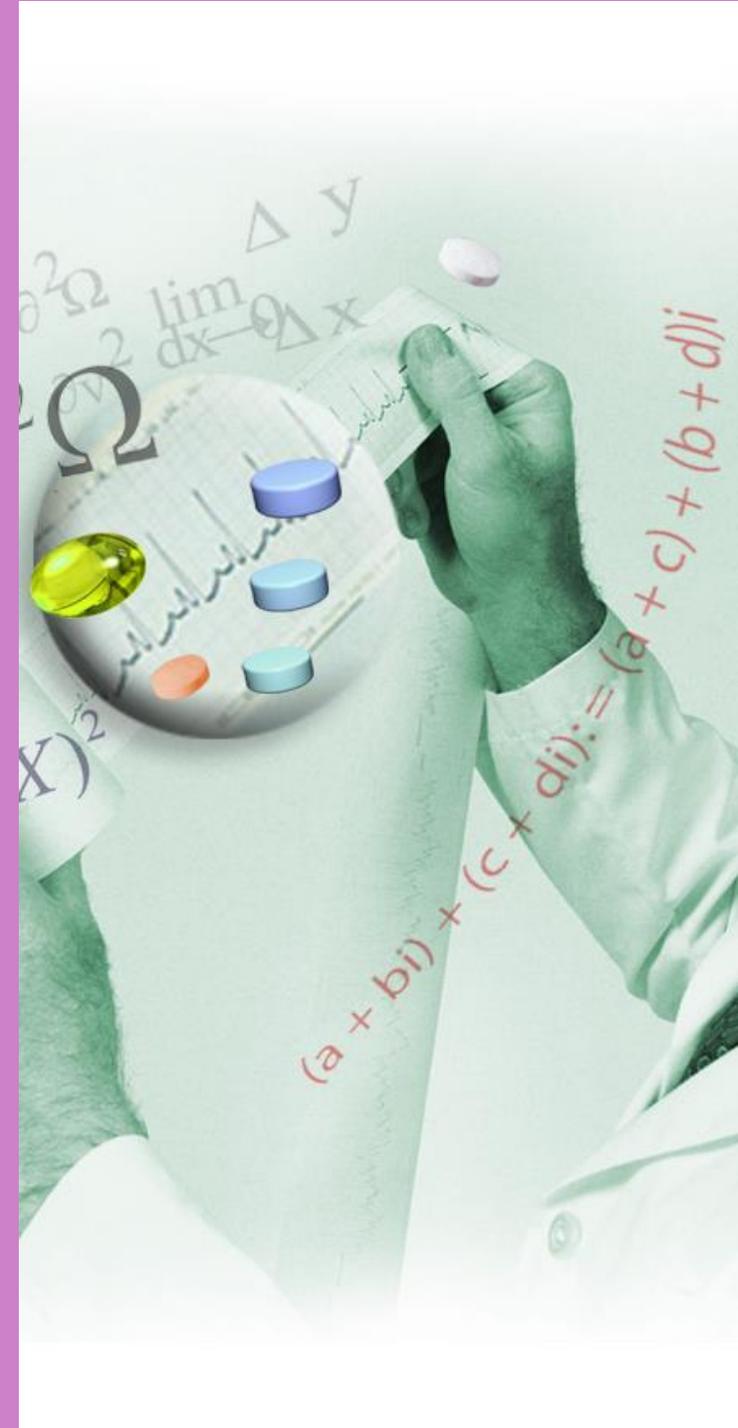
**PART 3:** Use of the conditional distribution to enhance genetic covariates analysis

To assess a PK **phenotype enrichment approach**

GENERAL METHODS

---

# SIMULATION STUDY ASSOCIATION TESTS



# GENERAL METHODS

# SIMULATION STUDY

# GENOTYPES

176 SNPs simulated based on the DNA microarray developed by Servier, retaining correlations between variants found in the human genome using a reference panel of Hapmap genotypes data set<sup>1</sup> and a specialised software (Hapgen2)<sup>2</sup>

1. International HapMap Consortium 2003

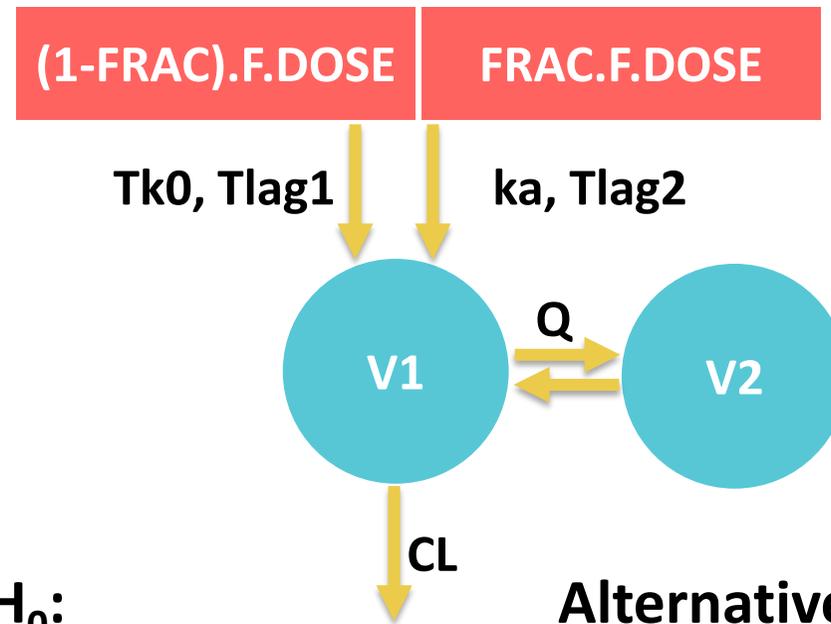
2. Su et al. 2011

# GENERAL METHODS

# SIMULATION STUDY

# POPULATION PHARMACOKINETIC MODEL

Nonlinearity on drug absorption (F and FRAC parameters)



**Null hypothesis  $H_0$ :**  
No genetic effect

**Alternative hypothesis  $H_1$ :**  
Genetic variants affect drug clearance

# GENERAL METHODS

# SIMULATION STUDY

# GENETIC EFFECT

Under  $H_1$ , 6 SNPs drawn randomly affect the clearance:

$$\log(CL_i) = \log(\mu_{CL}) + \sum_{k=1}^6 \beta_k \times SNP_{ik} + \eta_{i_{CL}}$$

$SNP_{ik} = \{0, 1, 2\}$ : additive genetic model

$\beta_k$ : effect size associated to the genotype  $SNP_{ik}$ , depends on:

$p_k$ : the frequency of the minor allele

$R_{GCk}$ : % of the interindividual variability in CL explained by the SNP<sup>1</sup>

SNP	1	2	3	4	5	6
$R_{GCk}$	1 %	2 %	3 %	5 %	7 %	12 %

# SIMULATION STUDY

## PHENOTYPES: NCA VS NLMEM

Individual PK profiles were simulated from PK model with genetic effects

- AUC estimated through **NCA**
  - normalised by the doses
- Individual clearances ( $EBE_{CL}$ ) estimated by **NonLinear Mixed effects Models (NLMEM)** under  $H_0$ 
  - Stochastic Approximation Expectation Maximisation algorithm (SAEM)<sup>1</sup>
  - Monolix software (v 4.2.2)<sup>2</sup>

All phenotypes were log-transformed

1. Kuhn and Lavielle. 2004

2. [www.lixoft.eu](http://www.lixoft.eu)

### Posterior PG analysis in a PK study

No assumption of genetic mechanisms influencing drug PK

The 176 simulated SNPs are tested

### Correlations between variants

### Family Wise Error Rate (FWER) correction

FWER controlled around 20%

FWER correction:  $1 - (1 - FWER)^{1/N_{SNP}}$

# GENERAL METHODS

# ASSOCIATION TESTS

## USUAL STRATEGY

### 1. Stepwise procedure

Iterative method

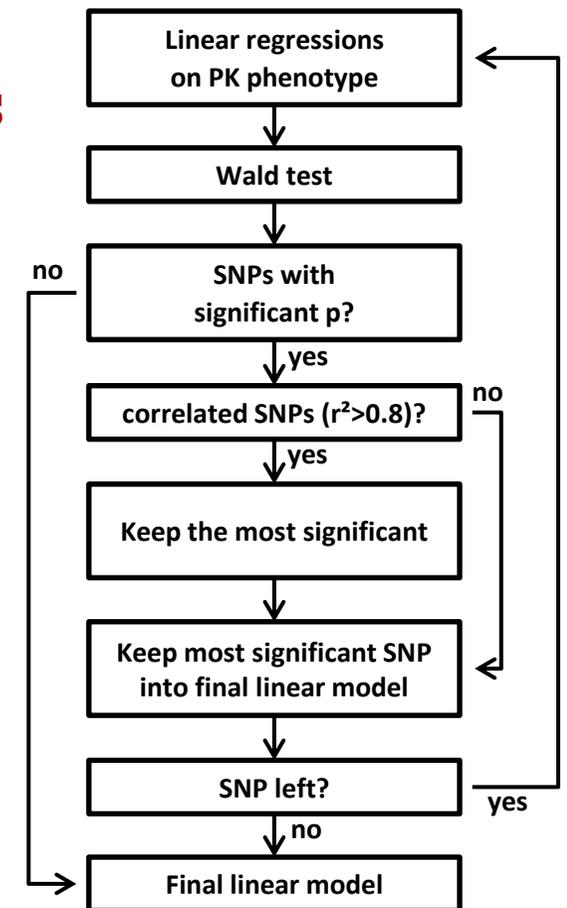
Effect sizes estimated through **univariate regressions**

$$\hat{\beta}_k = \operatorname{argmin}(Y - X\beta)^2$$

Variants selected using a Wald test

FWER correction for significance threshold

Taking into account correlations between variants<sup>1</sup>



1. Inspired from Lehr et al. 2010

# GENERAL METHODS

# ASSOCIATION TESTS

# PENALISED REGRESSIONS

## Multivariate analysis

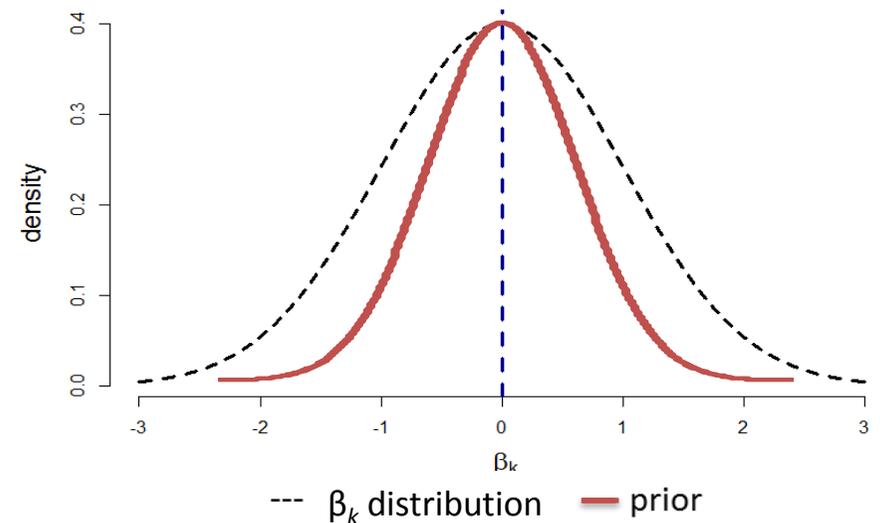
$$\hat{\beta}_k = \operatorname{argmin}(Y - X\beta)^2 + \textit{penalty}$$

## 2. Ridge regression<sup>1</sup>

Gaussian prior

Wald test for variable selection

FWER correction for significance threshold



# GENERAL METHODS

# ASSOCIATION TESTS

# PENALISED REGRESSIONS

## Multivariate analysis

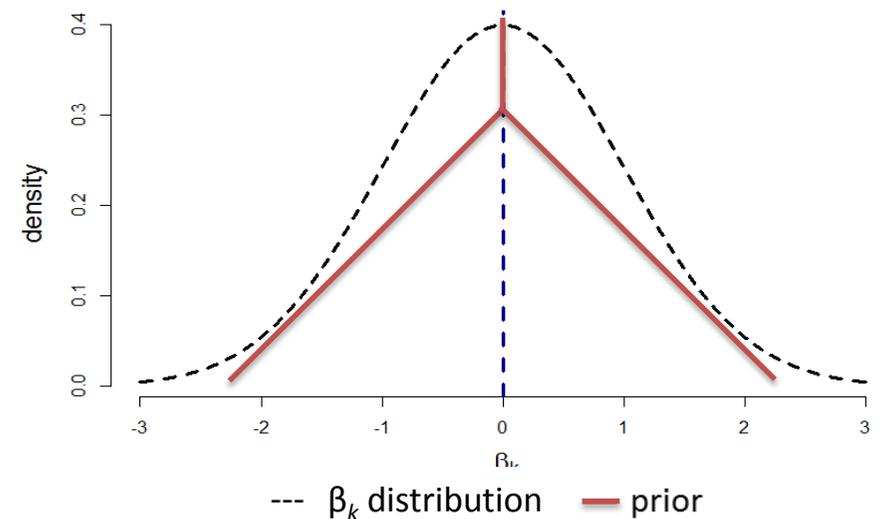
$$\hat{\beta}_k = \operatorname{argmin}(Y - X\beta)^2 + \textit{penalty}$$

### 3. Lasso<sup>1</sup>

Double Exponential (DE) prior

Set some coefficients to 0 (no test)

FWER correction to compute penalty



# GENERAL METHODS

# ASSOCIATION TESTS

# PENALISED REGRESSIONS

## Multivariate analysis

$$\hat{\beta}_k = \operatorname{argmin}(Y - X\beta)^2 + \textit{penalty}$$

### 4. HyperLasso<sup>1</sup>

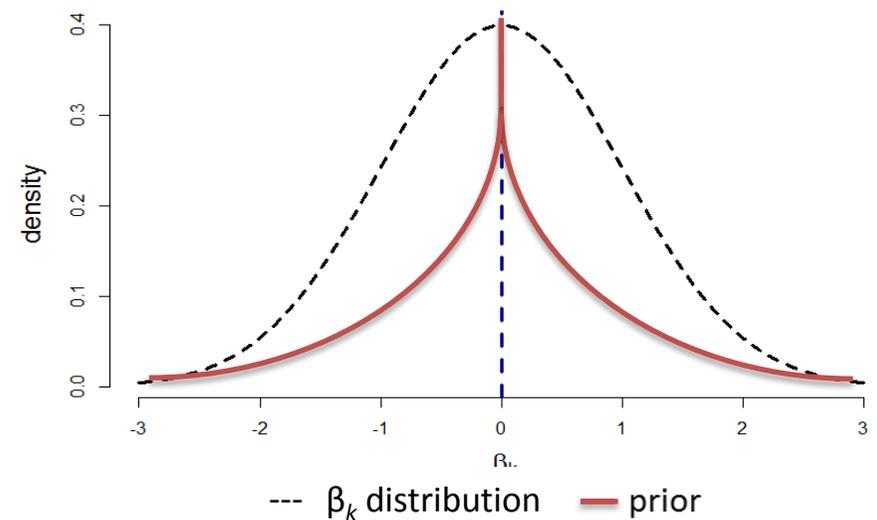
Normal exponential gamma (NEG) prior

shape and scale parameters

sparser solutions

Set some coefficients to 0 (no test)

FWER correction to compute penalty



PART 1

---

# ASSESSMENT OF PHARMACOGENETIC ANALYSIS METHODS



**S<sub>real</sub>**

## Phase I design

---

<b>Administration</b>	Single dose
<b>Number of subjects (<math>N_1</math>)</b>	78 subjects
<b>Elementary design (<math>\xi_{ij}</math>)</b>	16 sampling times <sup>a</sup>

---

<sup>a</sup> from 0.5 to 192h

**S<sub>large</sub>**

## Asymptotic conditions for N

---

<b>Administration</b>	Single dose
<b>Number of subjects (<math>N_1</math>)</b>	384 subjects
<b>Elementary design (<math>\xi_{ij}</math>)</b>	16 sampling times <sup>a</sup>

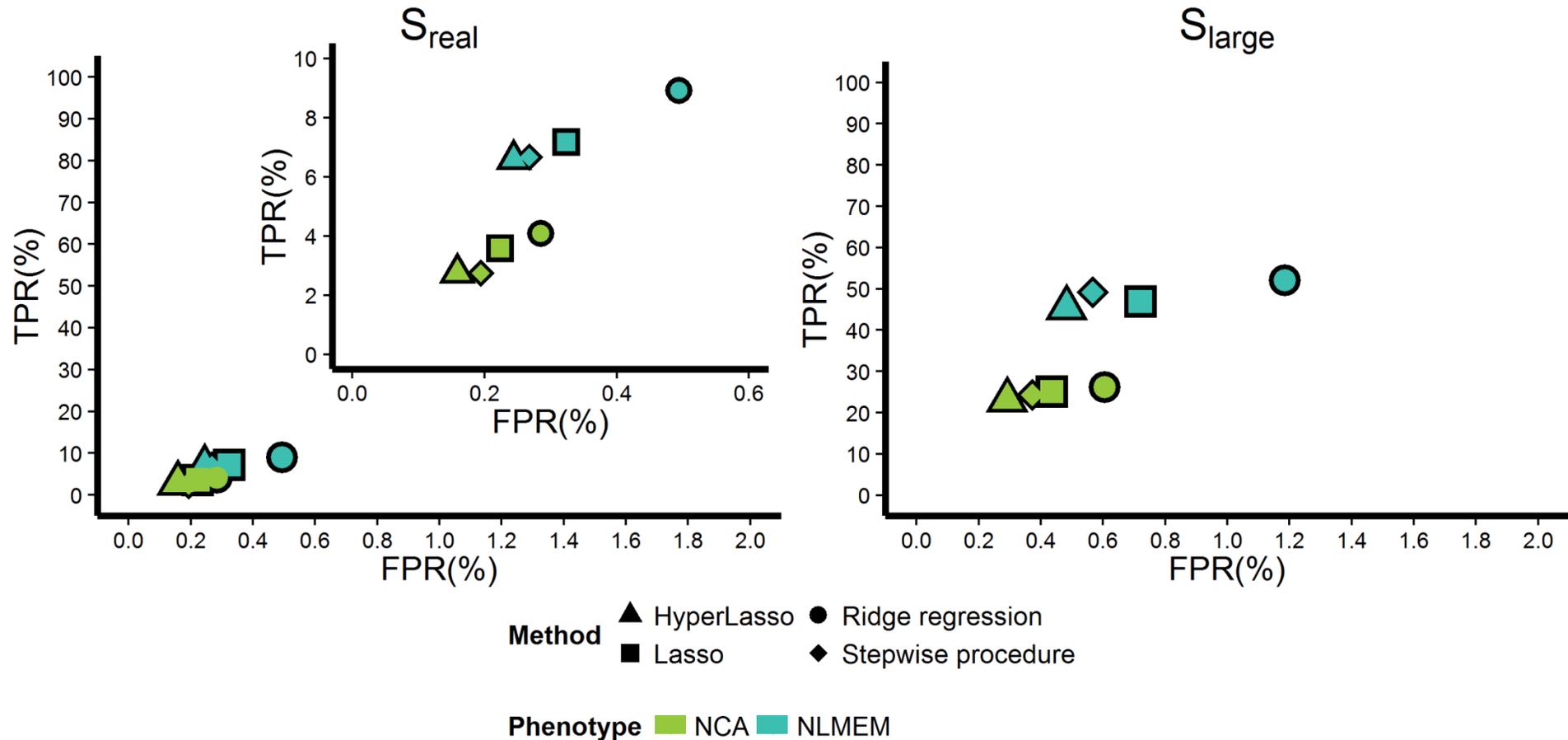
---

200 data sets were simulated for each scenario

# PART 1: RESULTS

# COMPARISON OF PHENOTYPE ESTIMATION METHODS

## NCA vs NLMEM



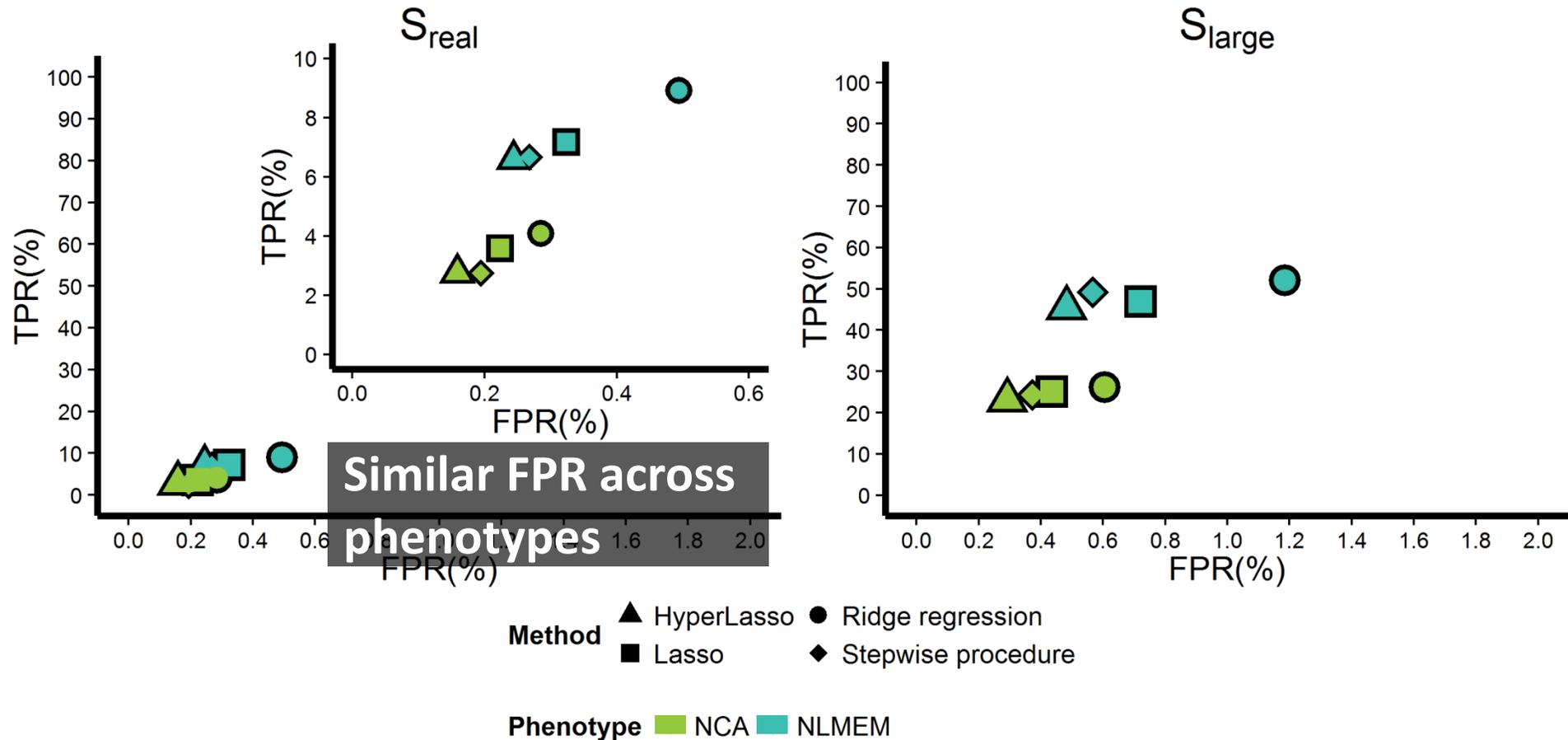
True positives (TP): SNP selected in the model and indeed associated to CL in the simulation

False positives (FP): SNP selected in the model but not present in the simulation

# PART 1: RESULTS

# COMPARISON OF PHENOTYPE ESTIMATION METHODS

## NCA vs NLMEM

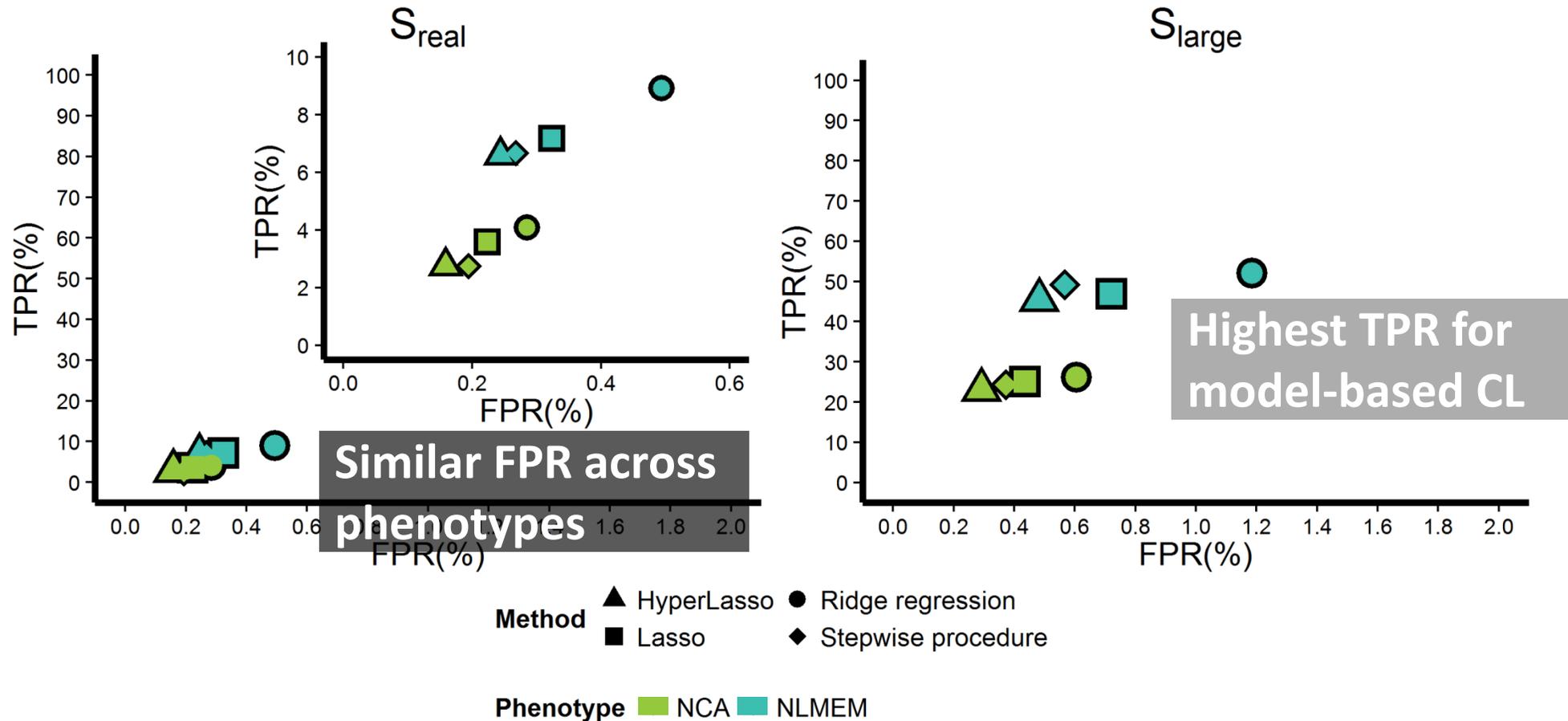


True positives (TP): SNP selected in the model and indeed associated to CL in the simulation

False positives (FP): SNP selected in the model but not present in the simulation

## COMPARISON OF PHENOTYPE ESTIMATION METHODS

## NCA vs NLMEM

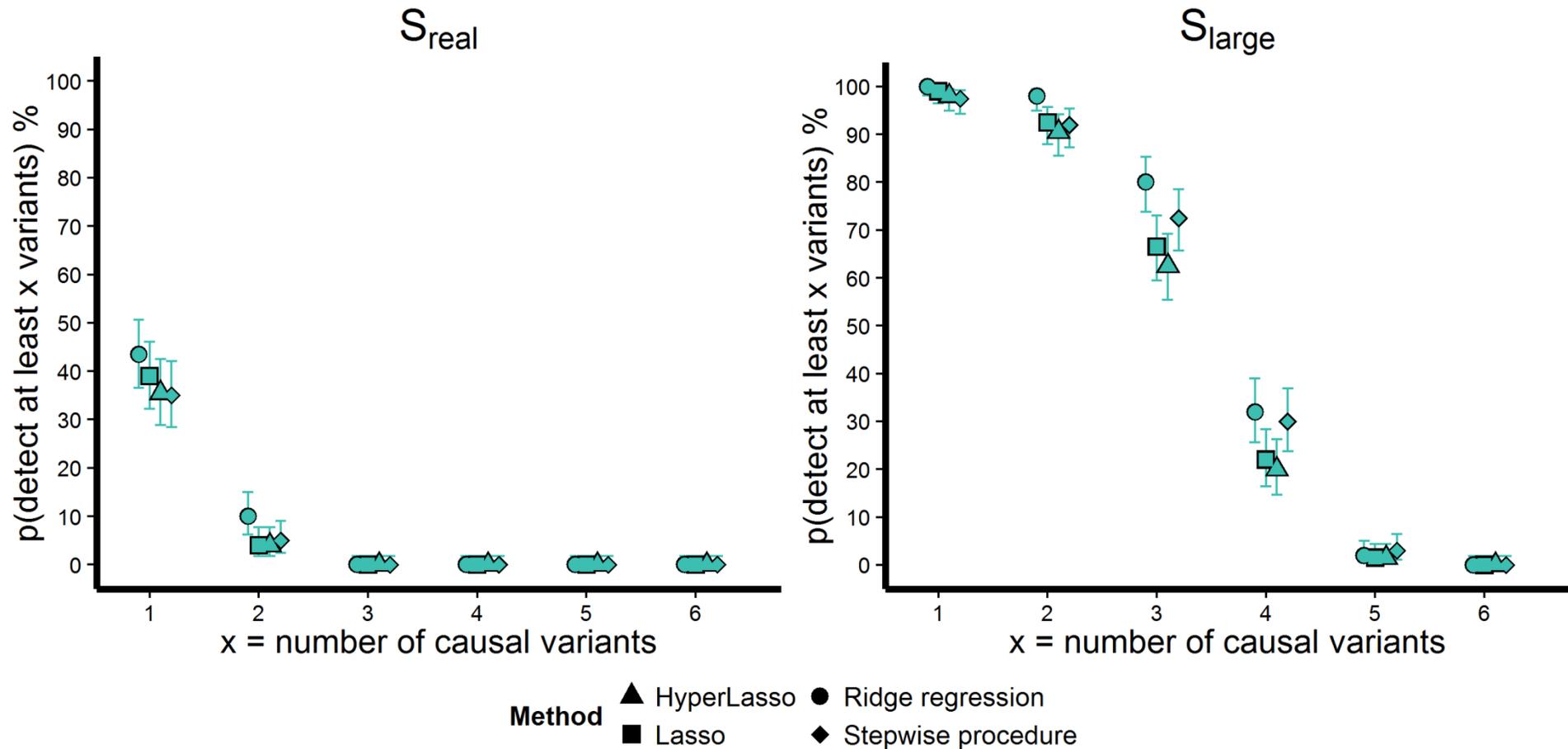


True positives (TP): SNP selected in the model and indeed associated to CL in the simulation

False positives (FP): SNP selected in the model but not present in the simulation

## COMPARISON OF ASSOCIATION TESTS

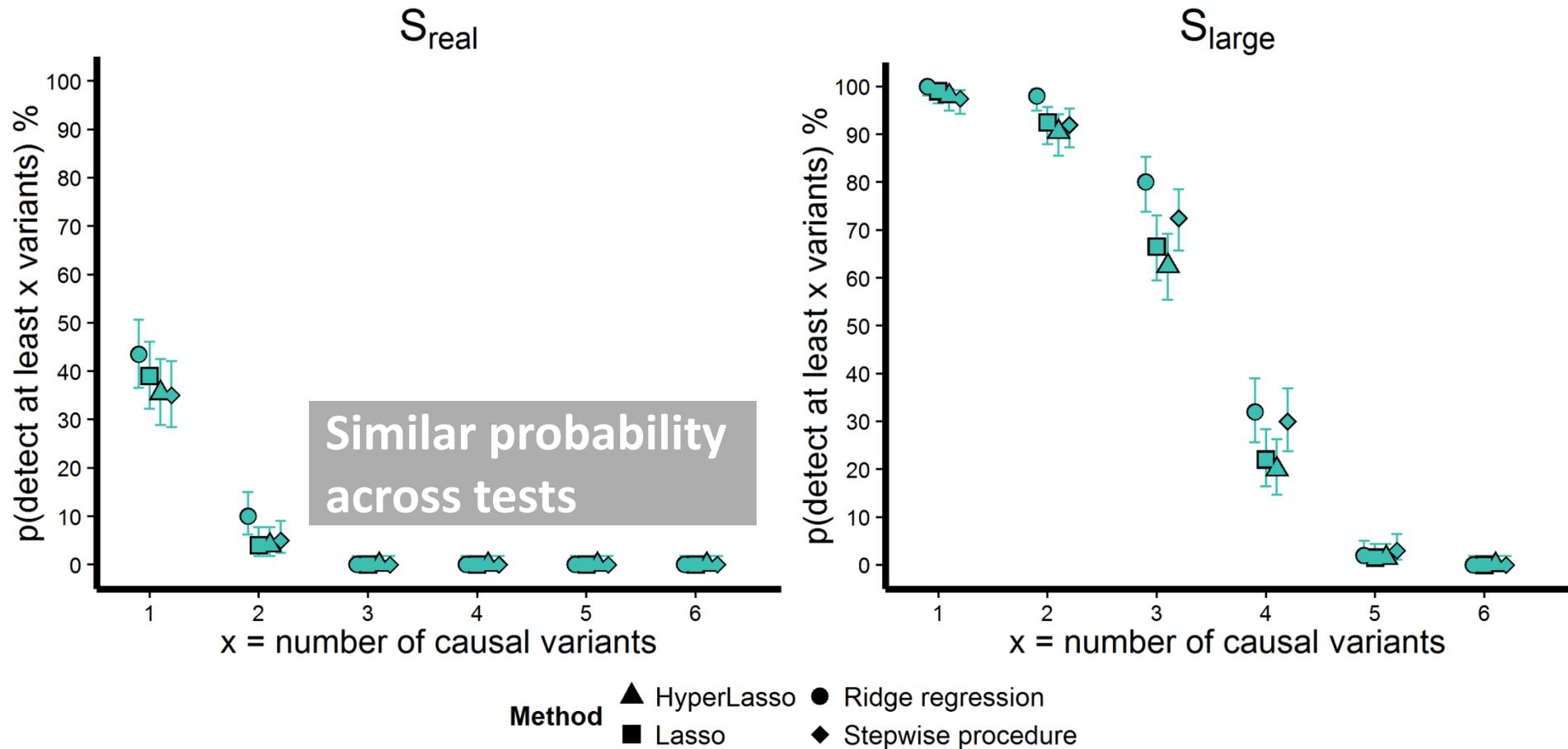
## PROBABILITY TO DETECT GENETIC EFFECTS ON NLMEM PHENOTYPE



Probability to detect genetic effects: % of data sets simulated where at least  $x$  SNPs ( $x=1, \dots, 6$ ) of the 6 SNPs are selected (200 simulated data set)

## COMPARISON OF ASSOCIATION TESTS

## PROBABILITY TO DETECT GENETIC EFFECTS ON NLMEM PHENOTYPE



Probability to detect genetic effects: % of data sets simulated where at least  $x$  SNPs ( $x=1, \dots, 6$ ) of the 6 SNPs are selected (200 simulated data set)

# PART 1: CONCLUSIONS

With a nonlinear PK, higher probability to detect genetic effects with a phenotype estimated through NLMEM

Similar power for the 4 association tests (penalised regressions or the stepwise procedure)

PART 2

---

# ENHANCE DETECTION OF GENETIC VARIANTS THROUGH COMBINED DESIGNS



PHOTOGRAPH BY ADAM VOORHES

# PART 2: METHODS

## SCENARIOS

Three phase II designs:

$N_2 = 306$  subjects at steady-state

	$PII_{3s.96h}^a$	$PII_{3s.24h}^a$	$PII_{1s.24h}$
Elementary design ( $\xi_{ij}$ )	2, 22, 96h	2, 22, 24h	24h

<sup>a</sup>designs optimised using PFIM<sup>1,2</sup>

Four analysis scenarios:

**SPI**: Phase I data only ( $N_1 = 78$ )

**SPI/PII<sub>3s.96h</sub>**: Phase I + Phase II data {2, 22, 96h}

**SPI/PII<sub>3s.24h</sub>**: Phase I + Phase II data {2, 22, 24h}

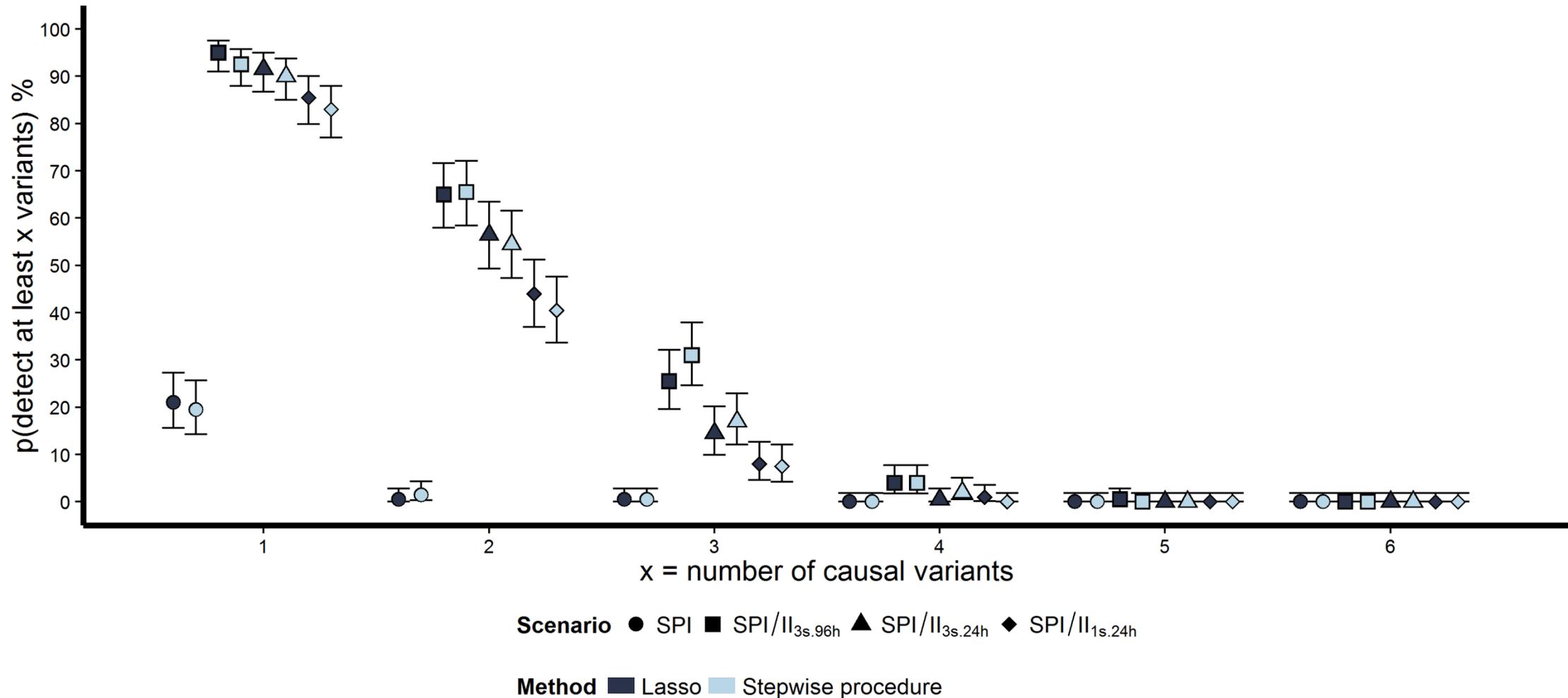
**SPI/PII<sub>1s.24h</sub>**: Phase I + Phase II data {24h}

200 data sets were simulated for each scenario

## PART2: RESULTS

# COMPARISON OF DESIGNS

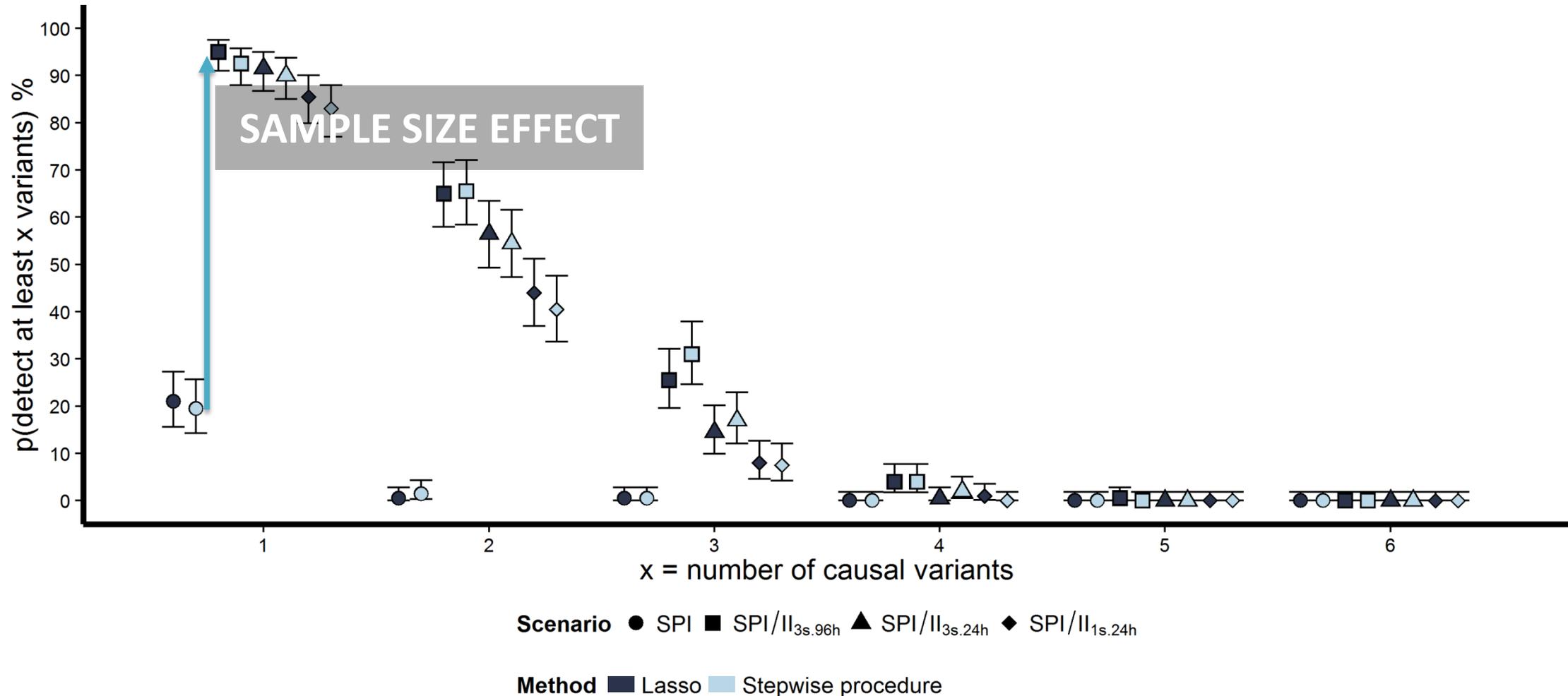
# PROBABILITY TO DETECT GENETIC EFFECTS ON NLMEM PHENOTYPE



## PART 2: RESULTS

# COMPARISON OF DESIGNS

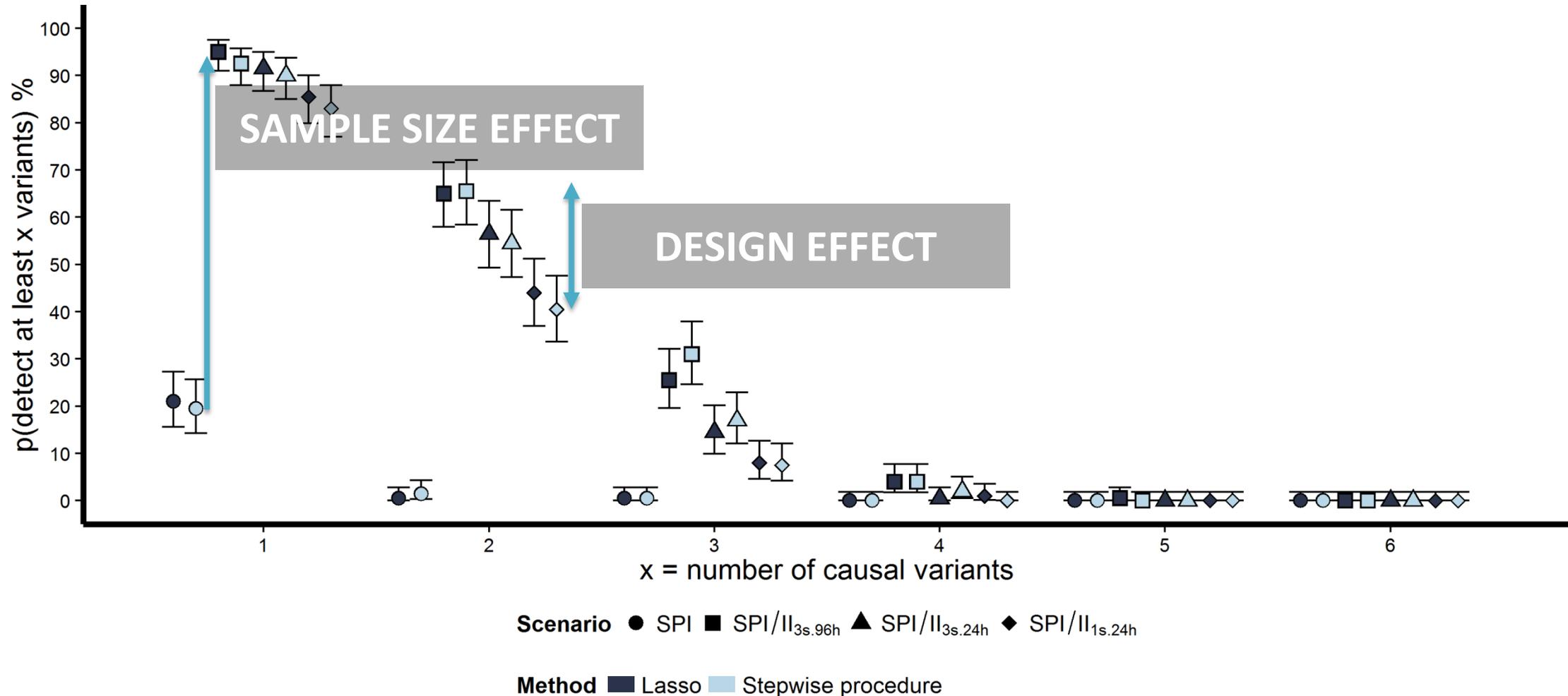
# PROBABILITY TO DETECT GENETIC EFFECTS ON NLMEM PHENOTYPE



## PART 2: RESULTS

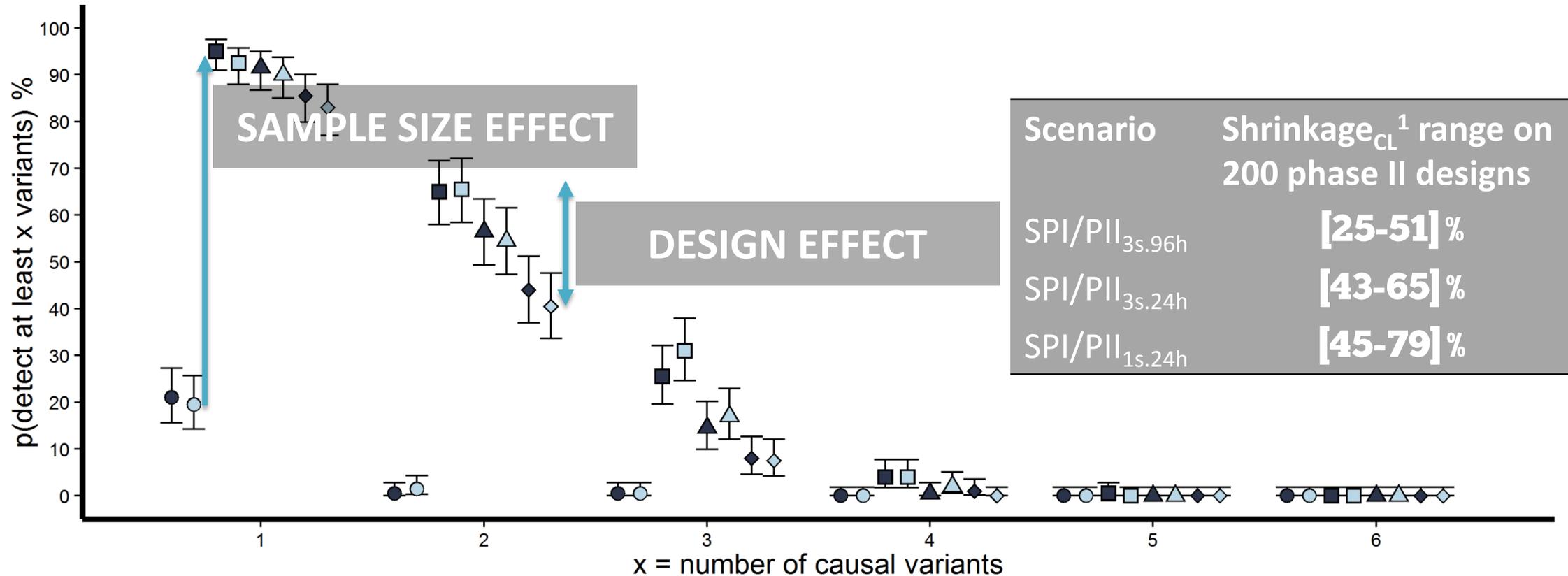
# COMPARISON OF DESIGNS

# PROBABILITY TO DETECT GENETIC EFFECTS ON NLMEM PHENOTYPE



# COMPARISON OF DESIGNS

## PROBABILITY TO DETECT GENETIC EFFECTS ON NLMEM PHENOTYPE



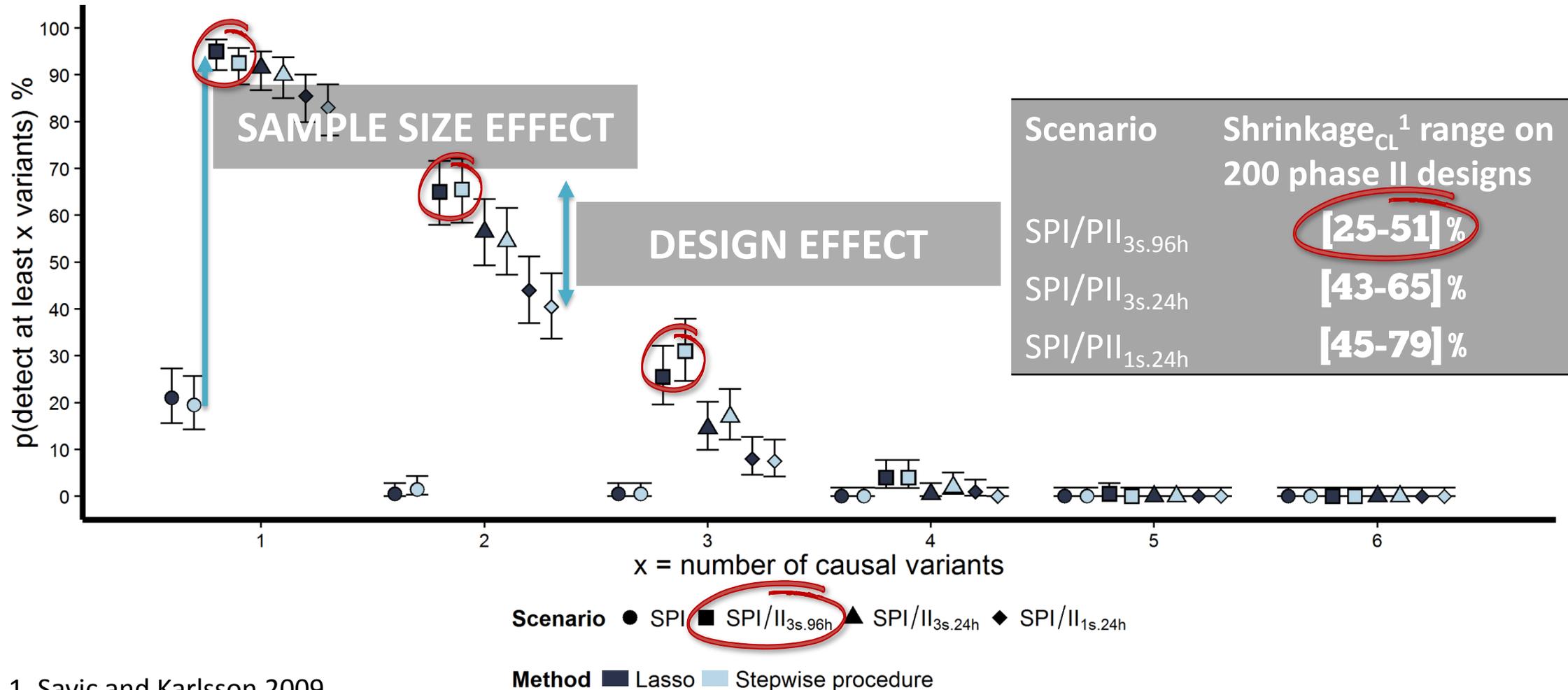
Scenario ● SPI ■ SPI/II<sub>3s.96h</sub> ▲ SPI/II<sub>3s.24h</sub> ◆ SPI/II<sub>1s.24h</sub>

Method ■ Lasso □ Stepwise procedure

# PART 2: RESULTS

## COMPARISON OF DESIGNS

### PROBABILITY TO DETECT GENETIC EFFECTS ON NLMEM PHENOTYPE



1. Savic and Karlsson 2009

## PART 2: CONCLUSIONS

Even with advanced methods (NLMEM, stepwise procedure or penalised regressions) results showed poor performance using phase I data only due to the small sample size

Significant improvement of the probability to detect realistic polymorphisms combining data from phase I and phase II in NLMEM

Mixing subsets with rich and sparse designs could also be performed within a phase II study

The design of phase II studies benefits from optimisation to a lower extent

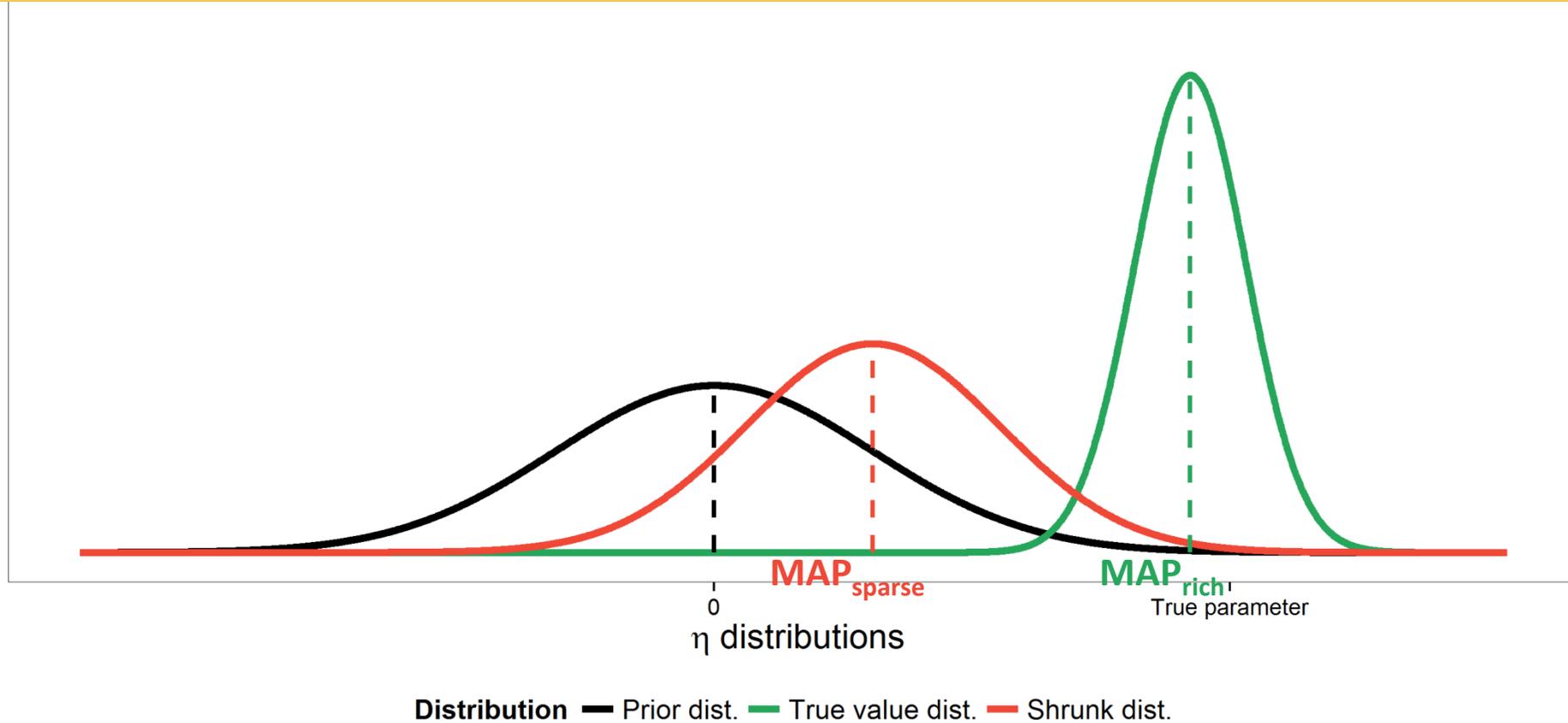
PART 3

---

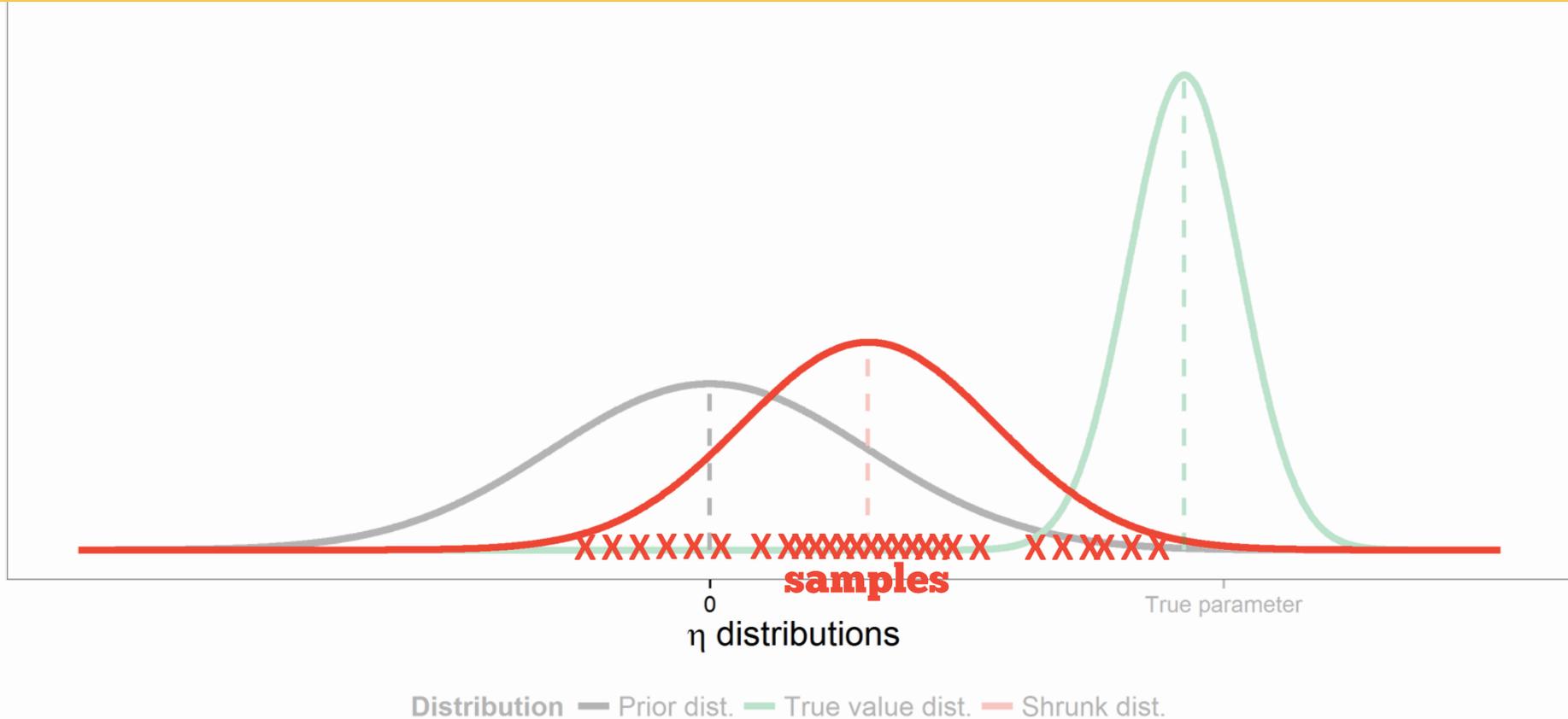
# USE OF THE CONDITIONAL DISTRIBUTION TO ENHANCE GENETIC COVARIATES ANALYSIS



# BAYESIAN APPROACH FOR INDIVIDUAL ESTIMATIONS



# BAYESIAN APPROACH FOR INDIVIDUAL ESTIMATIONS



Samples from conditional distribution computed using Monolix software<sup>1</sup>  
Proposed in diagnostic plots in last versions

# SAMPLES IN THE CONDITIONAL DISTRIBUTION SCENARIOS

Individual PK profiles were simulated from PK model with genetic effects

**1 SNP** drawn randomly to affect the clearance:

$$R_{GC} = \{5, 12, 20\%\}$$

N = 78 subjects

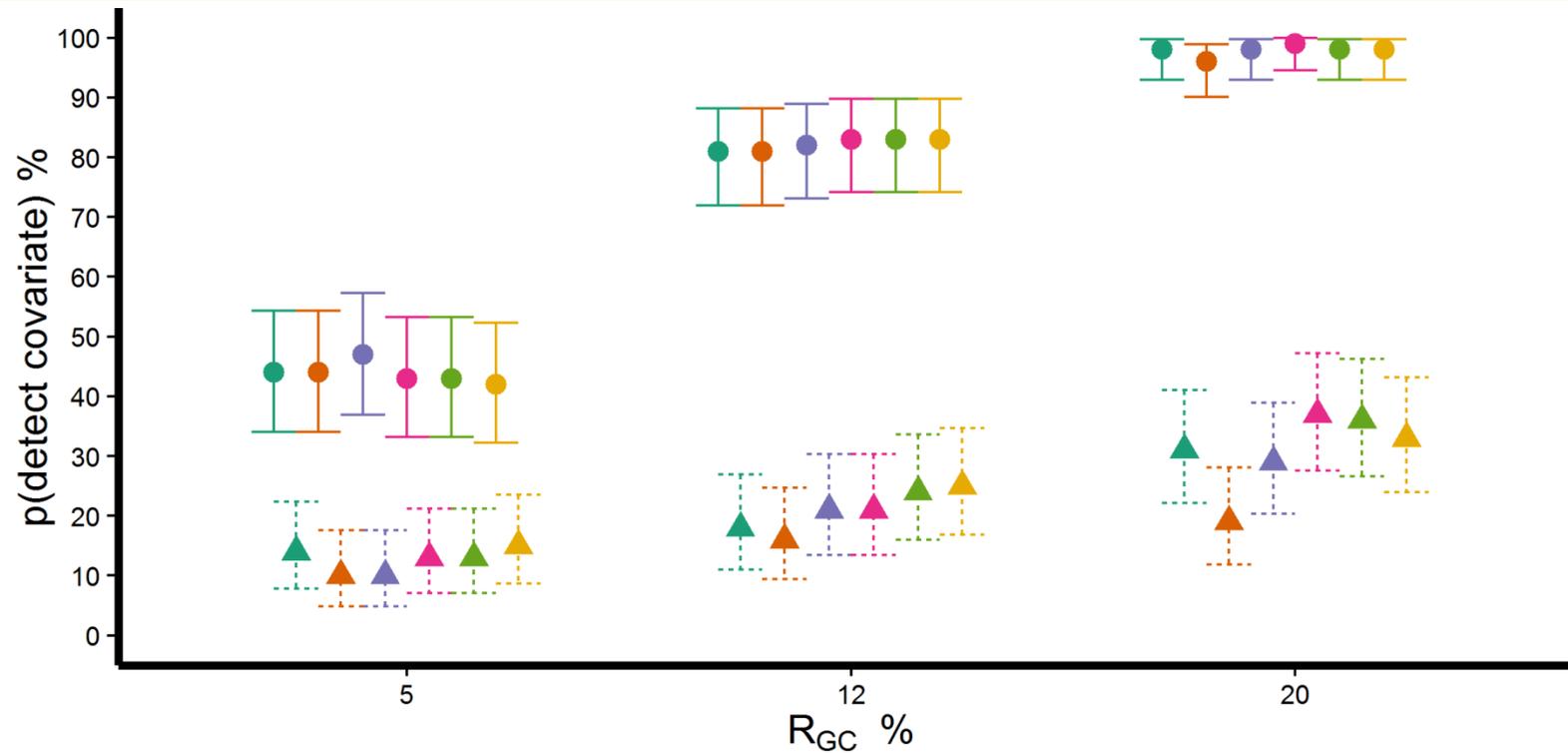
2 scenarios: Rich design (n = 16) or Sparse design (n = 1 with 3 groups)

Univariate regression (lm) on **individual parameters**  $\hat{\eta}_{CL_i}$

Linear mixed effects model (lme) on **samples from conditional distribution**

- random effect to handle correlations between the different samples for the same subject
- with different numbers of samples (from 3 to 600)

## PROBABILITY TO DETECT THE GENETIC EFFECT

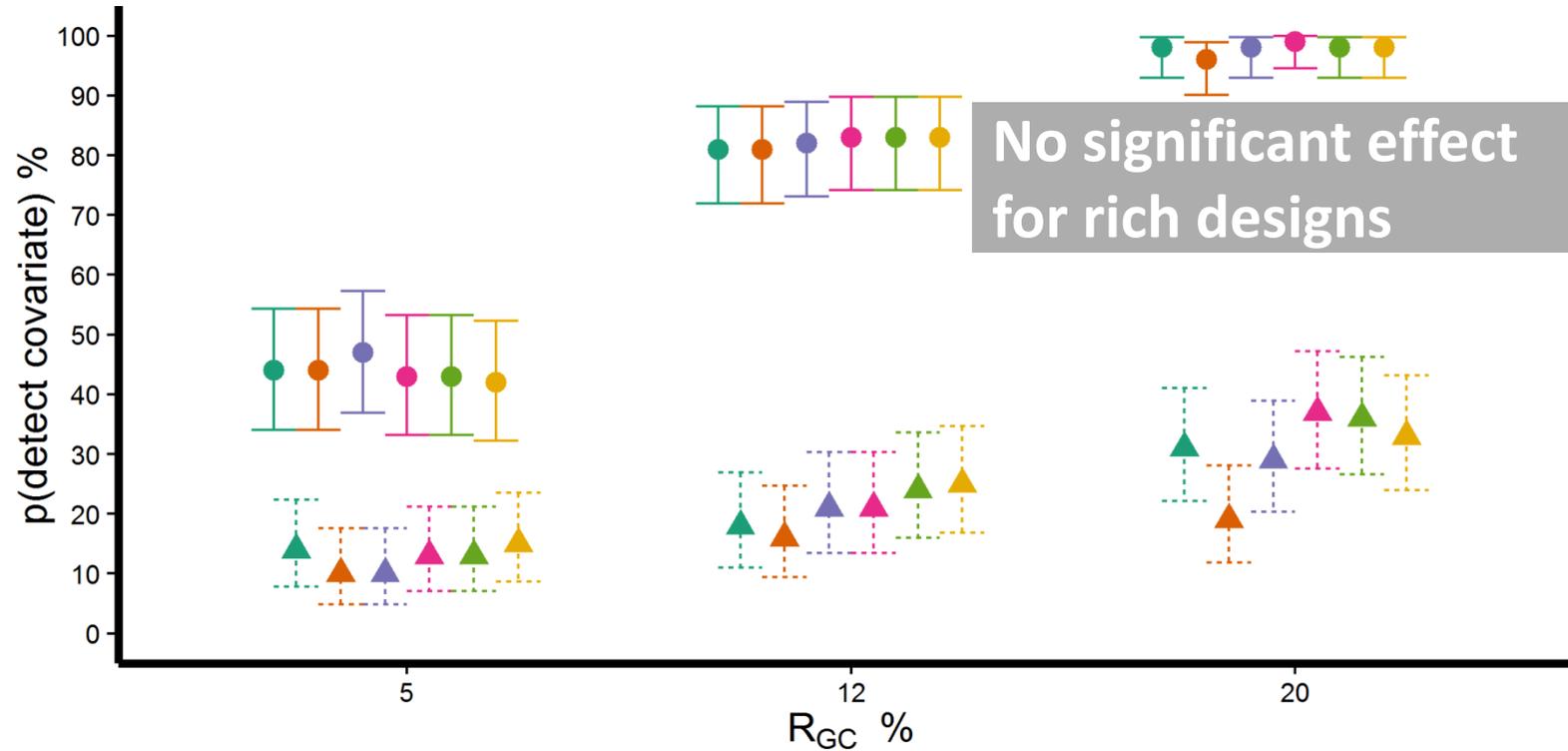


**Model** ■  $\hat{\eta}_{CL_i}$  (lme) ■ 10 samples (lme) ■ 300 samples (lme)  
■ 3 samples (lme) ■ 100 samples (lme) ■ 600 samples (lme)

**Design** ● Rich ▲ Sparse

**Design** — Rich ---- Sparse

## PROBABILITY TO DETECT THE GENETIC EFFECT

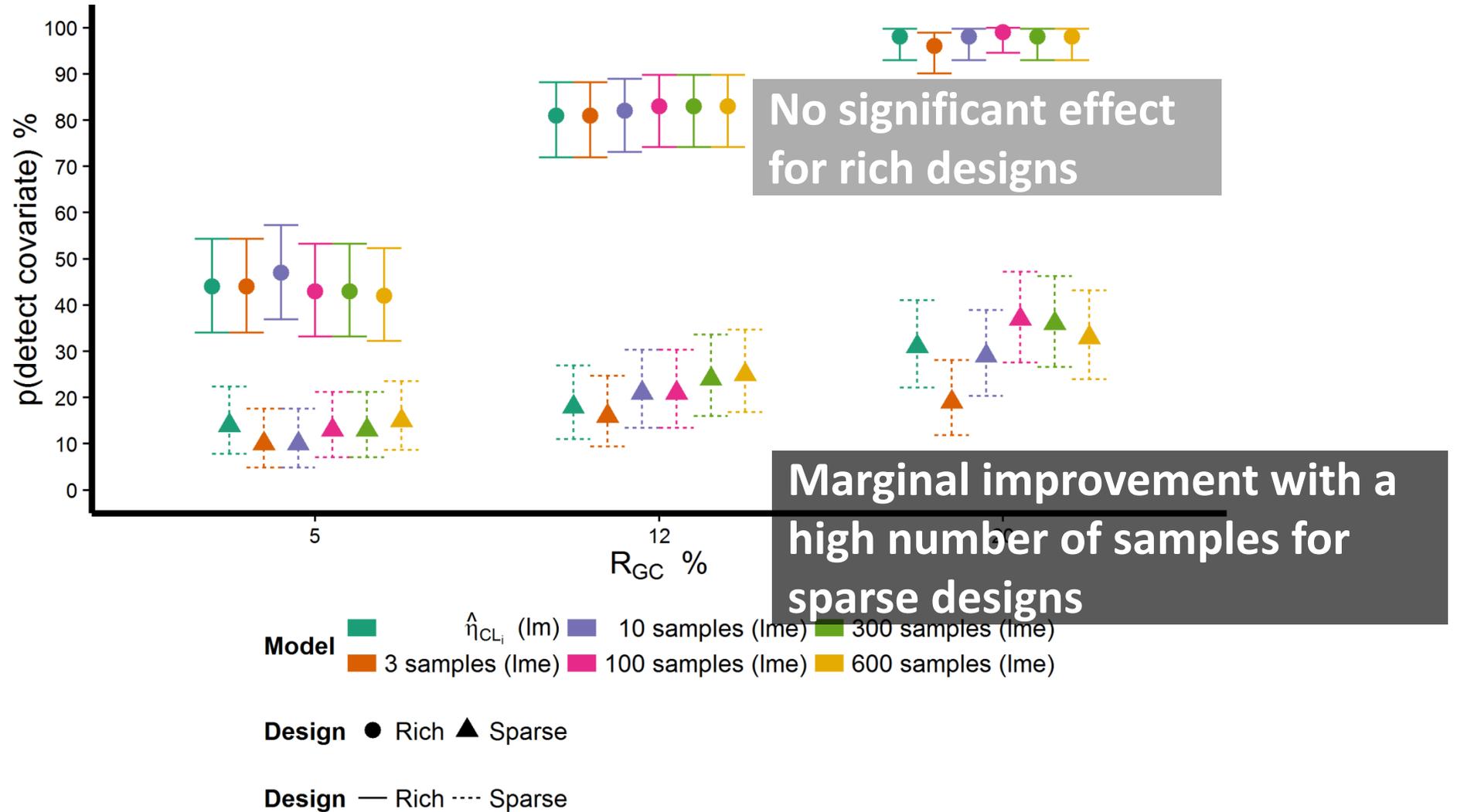


**Model**  $\hat{\eta}_{CL_i}$  (Im) 10 samples (Ime) 300 samples (Ime)  
 3 samples (Ime) 100 samples (Ime) 600 samples (Ime)

**Design** ● Rich ▲ Sparse

**Design** — Rich ---- Sparse

# PROBABILITY TO DETECT THE GENETIC EFFECT



# PART 3: CONCLUSIONS

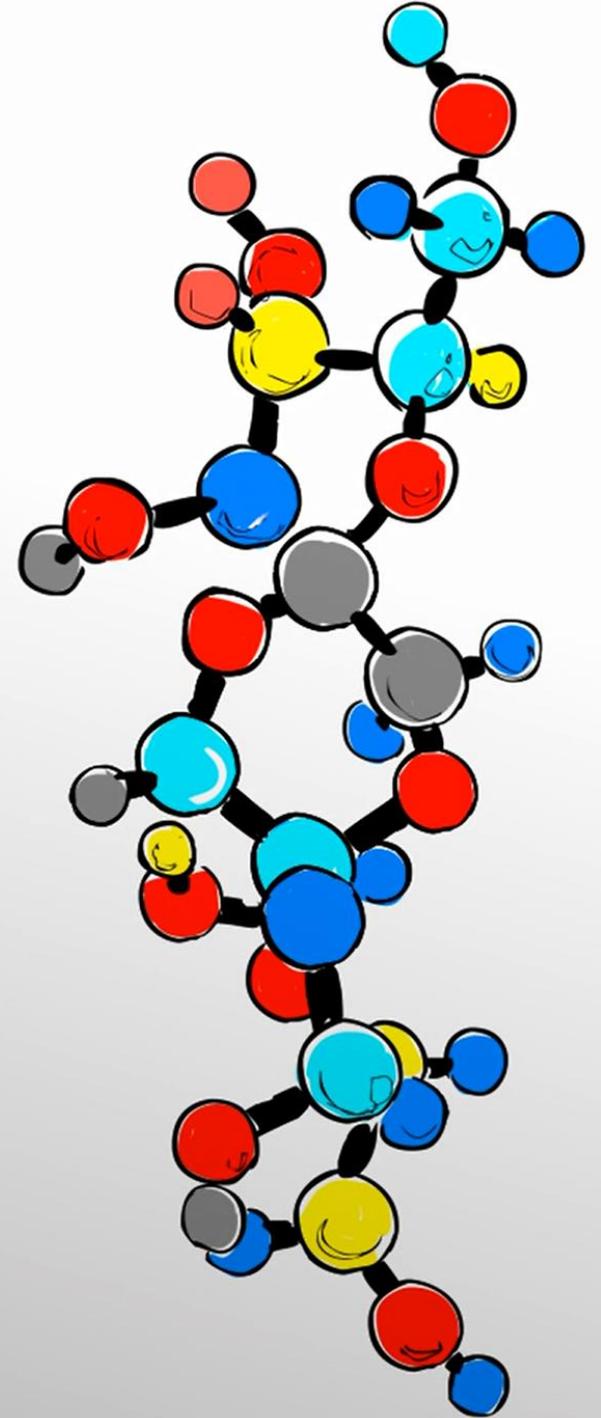
No benefit of using samples from the conditional distribution with rich designs

MAP from posterior distribution close to the true parameter

Marginal improvement in the probability to detect the genetic variant for sparse designs requires a large number of samples

---

# CONCLUSION



# RECOMMENDATIONS FOR PHARMACOGENETIC ANALYSIS

## **Phenotypes**

Modelling approaches should be preferred to estimate the PK phenotype

## **Association methods**

No benefit of penalised regressions

## **Design**

We recommend

to combine analysis of phase I and phase II data for the exploration of genetic associations

to prospectively optimise the phase II study design

# LIMITS AND PERSPECTIVES

## Limits

Only one setting investigated in simulations  
additive genetic model  
nonlinear PK

No re estimation of EBE conditional to the genetic variants in stepwise procedure

## Perspectives

Develop joint estimation/selection with penalised regression in NLMEM<sup>1</sup>

Investigate effect of model misspecifications on detection probability  
correlation between model parameters

1. Bertrand et al. PAGE 2013

AS ALWAYS  
**THANKS FOR YOUR ATTENTION**

# Acknowledgments



**Emmanuelle Comets**



**Marylore Chenel**



**Julie Bertrand**

**Servier Clinical PK & PMx team**

**Bernard Walther  
Laurent Ripoll**



**IAME team**

**France Mentré**

**PAGE committee**



We are grateful to Marc Lavielle for help implementing the imputations in the conditional distribution